

The Chinese University Of Hong Kong  
Joint Graduate Student Seminar  
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# Quorum Sensing in Yeast

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# Contents

- Introduction to quorum sensing
- Quorum sensing in yeast
  - Farnesol
  - Tyrosol
  - Other QSMs found in yeast
- Application of QSMs
- Conclusion
- References

# Introduction to quorum sensing

## Discovery of quorum sensing:

- Marine bioluminescent bacteria
- High cell concentration: luminesce  
Below threshold: do not luminesce
- Accumulation of secreted autoinducer signaling molecules  
→ Transcription of luciferase coding gene
- Similar phenomena and molecules in other bacteria

# Introduction to quorum sensing

Quorum sensing:

- Cell-to-cell communication
- Cell-density-dependent gene expression
- Accumulation of quorum sensing molecules(QSMs)
  - Reaching a critical threshold in concentration
  - Activation of specific signaling pathway

# Introduction to quorum sensing

## Criteria of quorum sensing molecules

- By Winzer and colleagues(2002)

### 1. Be produced

- During specific stages of growth
- Under certain physiological conditions
- In response to environmental changes

# Introduction to quorum sensing

## Criteria of quorum sensing molecules

- By Winzer and colleagues(2002)
2. Accumulate extracellularly & recognized by specific receptor
  3. Trigger concerted response when concentration reaches threshold
  4. Responses beyond metabolization or detoxification

# Introduction to quorum sensing

- Physiological reactions:

Biofilm formation

Bioluminescence

Virulence

Antibiotic secretion

Phenotype switch

etc.

**Efficient and effective usage of energy!**

# Quorum sensing phenomenon in yeast?

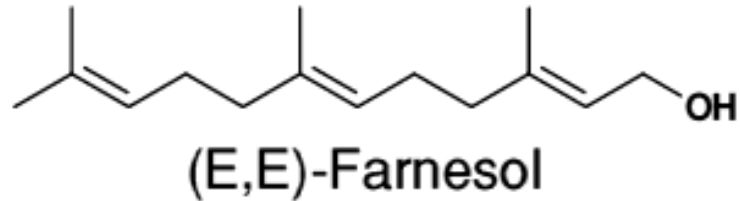


# Quorum sensing phenomenon in yeast?

Inoculum size effect

- Yeast-mycelium dimorphism
- Initial cell concentration  
→ Predomination of one morphology
- E.g. *Candida albicans*  
≥10<sup>6</sup> cells/ml: budding yeast  
<10<sup>6</sup> cells/ml: germ tubes and mycelia

# Discovery of farnesol



[http://wildflowerfinder.org.uk/Flowers/M/MockOrange/\(E\\_E\)Farnesol.png](http://wildflowerfinder.org.uk/Flowers/M/MockOrange/(E_E)Farnesol.png)

- Isolated by Hornby and colleagues(2001)
- First QSM identified in fungi
- Isolated from supernatant of *C. albicans* culture
- Inhibits filamentation

Control:  
% mycelia >>  
% budding  
yeast

TABLE 1. QSM cross-reactivity of A72 with five other strains of *C. albicans*

Cell type	Morphology results (mean % ± SD) <sup>a</sup> with:		
	37°C control	Supernatant from A-72 <sup>b</sup>	A-72 cells <sup>c</sup>
A-72	16 ± 1 (Y) 83 ± 1 (M)	71 ± 4 (Y) 26 ± 4 (M)	73 ± 4 (Y) 24 ± 3 (M)
MEN	49 ± 6 (Y) 47 ± 7 (M)	99 ± 1 (Y) 1 ± 1 (M)	61 ± 3 (Y) 37 ± 2 (M)
SG 5314	8 ± 1 (Y) 92 ± 1 (M)	44 ± 6 (Y) 53 ± 2 (M)	61 ± 3 (Y) 39 ± 3 (M)
LGH 1095	19 ± 2 (Y) 78 ± 3 (M)	88 ± 3 (Y) 10 ± 2 (M)	73 ± 1 (Y) 27 ± 1 (M)
SG 3314	20 ± 9 (Y) 80 ± 9 (M)	45 ± 9 (Y) 50 ± 10 (M)	74 ± 4 (Y) 21 ± 5 (M)
SG 10261	25 ± 7 (Y) 72 ± 8 (M)	57 ± 12 (Y) 34 ± 15 (M)	44 ± 9 (Y) 55 ± 10 (M)

+ Supernatant:  
% budding yeast ↑, % mycelia ↓

# Discovery of farnesol

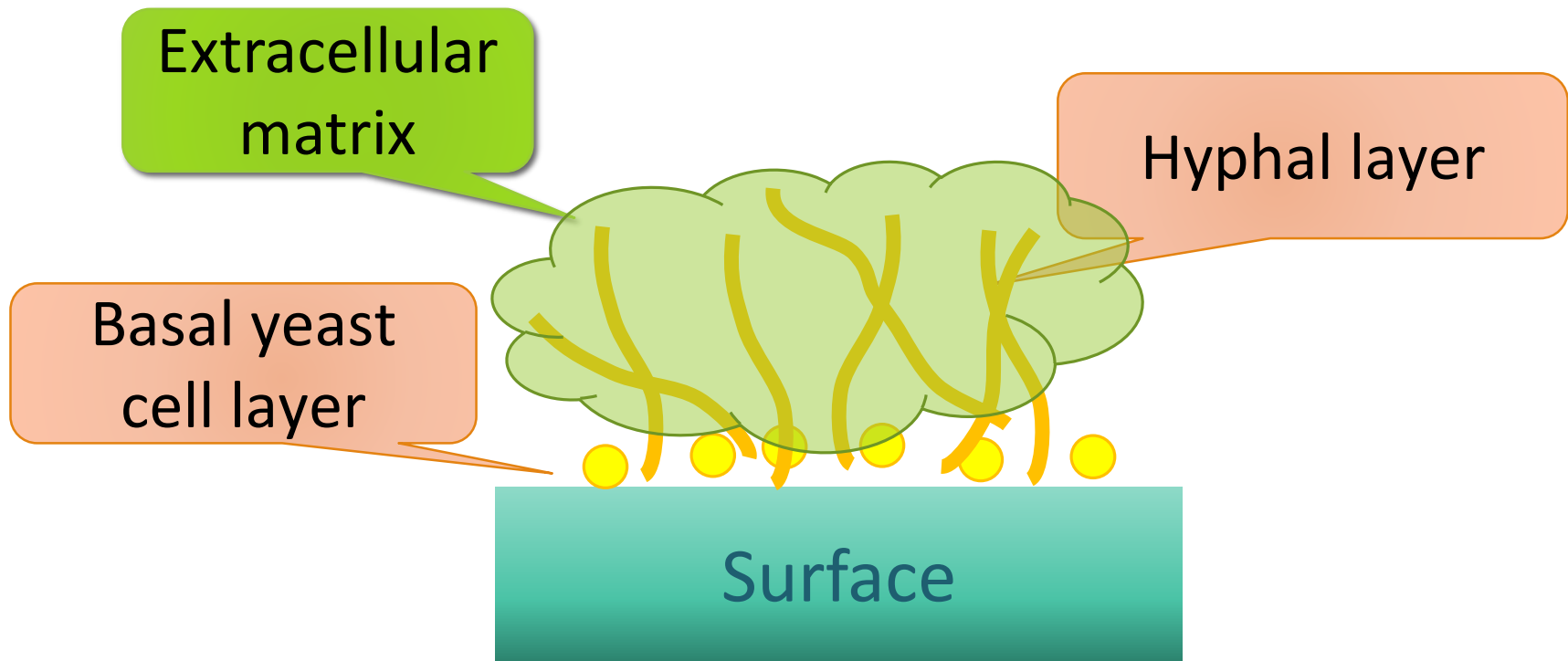
Other properties found by Hornby:

- Commercial farnesol has same effect
- Production:
  - Roughly proportional to cell density
  - Independent of growth media, carbon or nitrogen source
- No effects on growth rate
- Inhibit germ tube formation inducers:  
L-proline, N-acetylglucosamine and serum

# Other physiological effects of farnesol on *C. albicans*

Farnesol disturb formation of biofilm structure

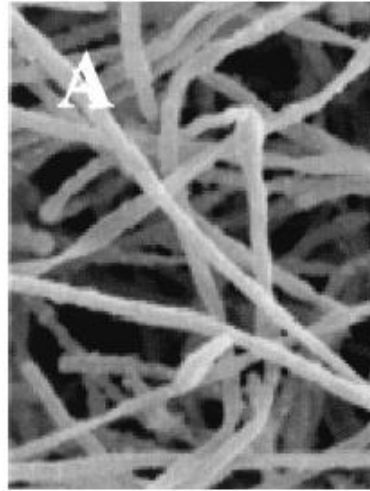
Mature *C. albicans* biofilm structure:



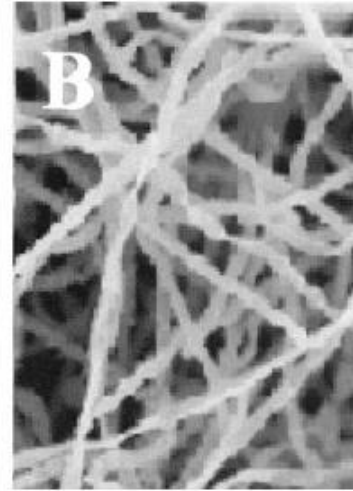
# Other physiological effects of farnesol on *C. albicans*

Farnesol disturb formation of biofilm structure

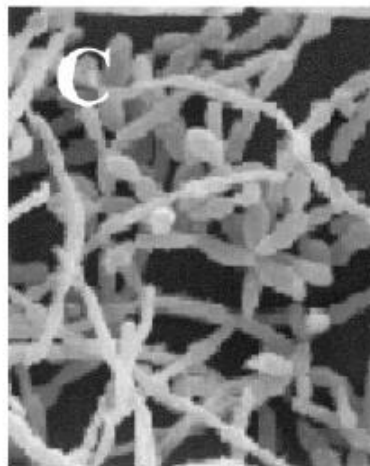
Control  
(w/o farnesol)



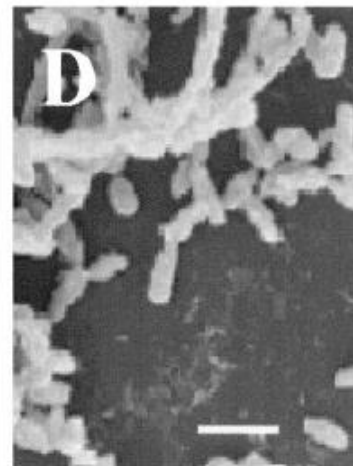
3 $\mu$ M  
Farnesol



30 $\mu$ M



300 $\mu$ M



Before  
filamentation,  
inhibition

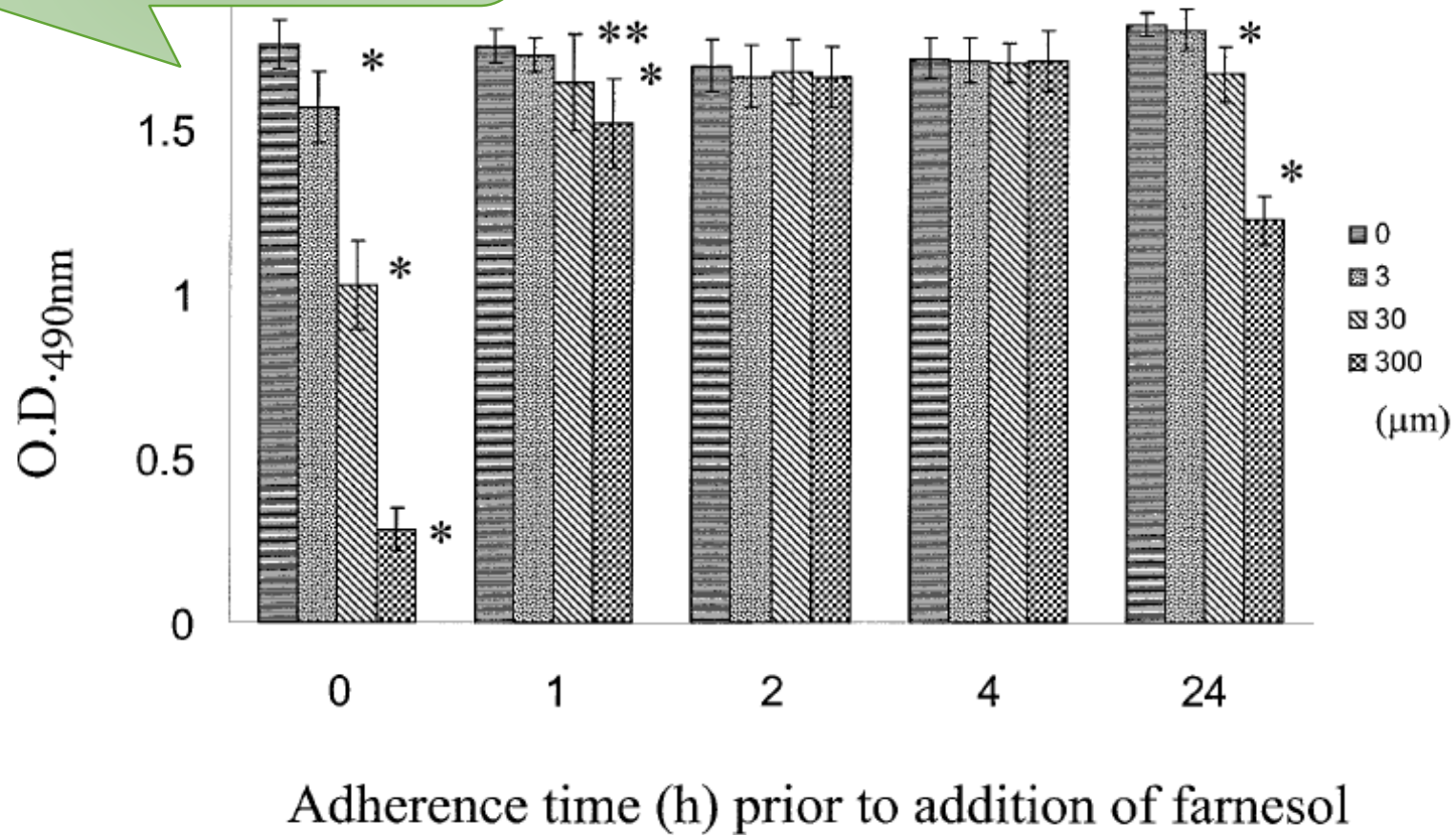


FIG. 1. Effect of farnesol on *C. albicans* biofilm formation. Different farnesol concentrations (0, 3, 30, and 300  $\mu\text{m}$ ) were added to *C. albicans* cells at different times after attachment (0, 1, 2, and 4 h), and the cells were incubated under biofilm-growing conditions; farnesol was also added to preformed (24-h) biofilms, which were incubated for an additional 24 h. The extent of biofilm formation was estimated by the XTT reduction assay. The values are mean absorbance values and standard deviations for 10 independent biofilms. Statistically significant differences (as determined by Student's *t* test, compared to biofilms formed in the absence of farnesol) are indicated as follows: one asterisk,  $P < 0.01$ ; two asterisks,  $P < 0.05$ . O.D.<sub>490nm</sub>, optical density at 490 nm.

After filamentation  
initiates, no effect

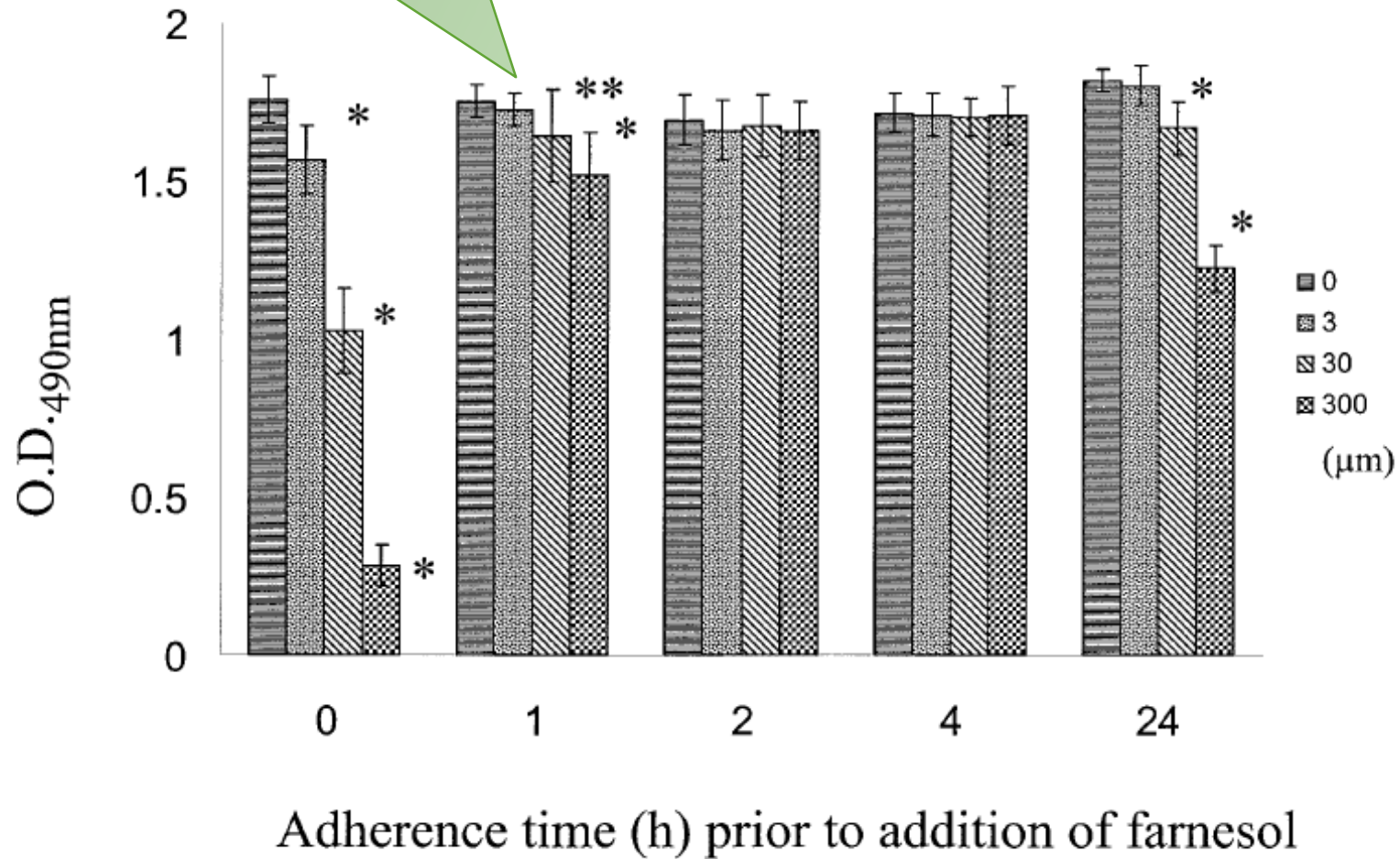


FIG. 1. Effect of farnesol on *C. albicans* biofilm formation. Different farnesol concentrations (0, 3, 30, and 300 μm) were added to *C. albicans* cells at different times after attachment (0, 1, 2, and 4 h), and the cells were incubated under biofilm-growing conditions; farnesol was also added to preformed (24-h) biofilms, which were incubated for an additional 24 h. The extent of biofilm formation was estimated by the XTT reduction assay. The values are mean absorbance values and standard deviations for 10 independent biofilms. Statistically significant differences (as determined by Student's *t* test, compared to biofilms formed in the absence of farnesol) are indicated as follows: one asterisk,  $P < 0.01$ ; two asterisks,  $P < 0.05$ . O.D.<sub>490nm</sub>, optical density at 490 nm.



New yeast cells: response to farnesol  
possibly related to biofilm dispersal

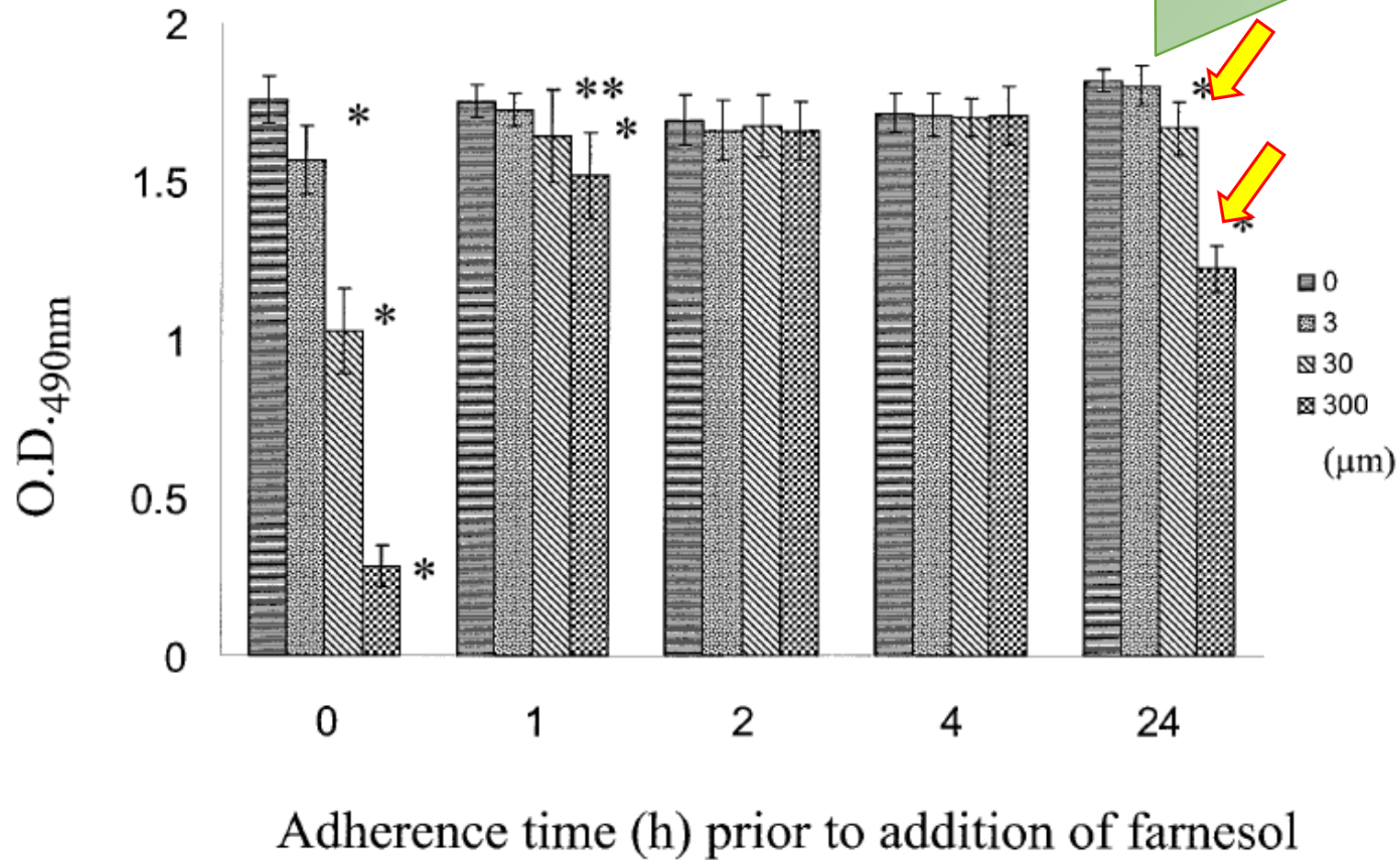


FIG. 1. Effect of farnesol on *C. albicans* biofilm formation. Different farnesol concentrations (0, 3, 30, and 300  $\mu\text{m}$ ) were added to *C. albicans* cells at different times after attachment (0, 1, 2, and 4 h), and the cells were incubated under biofilm-growing conditions; farnesol was also added to preformed (24-h) biofilms, which were incubated for an additional 24 h. The extent of biofilm formation was estimated by the XTT reduction assay. The values are mean absorbance values and standard deviations for 10 independent biofilms. Statistically significant differences (as determined by Student's *t* test, compared to biofilms formed in the absence of farnesol) are indicated as follows: one asterisk,  $P < 0.01$ ; two asterisks,  $P < 0.05$ . O.D.<sub>490nm</sub>, optical density at 490 nm.

# Other physiological effects of farnesol on *C. albicans*

## Oxidative stress

- Promote protection against oxidative stress

## Induction of apoptosis

## Modulation on drug efflux pump

- Specifically on ABC multidrug transporters
- Synergistic effect with azole & amphotericin B

# Physiological effects of farnesol on other microbes

Biofilm inhibition	Growth inhibition	Apoptosis	Cell death	Filamentation inhibition
<i>Candida dubliniensis</i>	<i>Candida parapsilosis</i>	<i>Aspergillus fumigatus</i>	<i>Saccharomyces cerevisiae</i>	<i>Candida dubliniensis</i>
<i>Candida parapsilosis</i>	<i>Acinetobacter baumannii</i>	<i>Aspergillus nidulans</i>	<i>Staphylococcus aureus</i>	<i>Aspergillus nidulans</i>
<i>Staphylococcus aureus</i>	<i>Aspergillus fumigatus</i>	<i>Fusarium graminearum</i>	<i>Staphylococcus epidermidis</i>	<i>Paracoccidiodes brasiliensis</i>
<i>Streptococcus mutans</i>	<i>Paracoccidiodes brasiliensis</i>			
	<i>Saccharomyces cerevisiae</i>			
	<i>Streptococcus mutans</i>			

# Another QSM from *C. albicans* -- Tyrosol

## Tyrosol is a quorum-sensing molecule in *Candida albicans*

Hao Chen\*, Masaki Fujita†, Qinghua Feng\*\*‡, Jon Clardy†, and Gerald R. Fink\*§

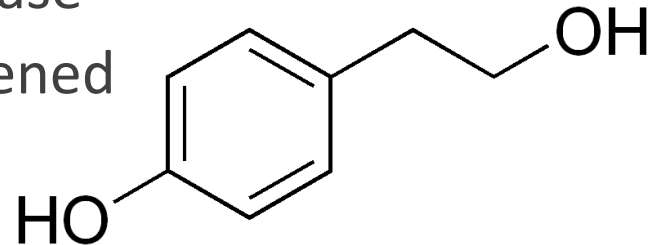
\*Whitehead Institute for Biomedical Research, Nine Cambridge Center, Cambridge, MA 02142; and †Department of Biological Chemistry and Molecular Pharmacology, Harvard Medical School, 240 Longwood Avenue, Boston, MA 02115

Contributed by Gerald R. Fink, February 27, 2004

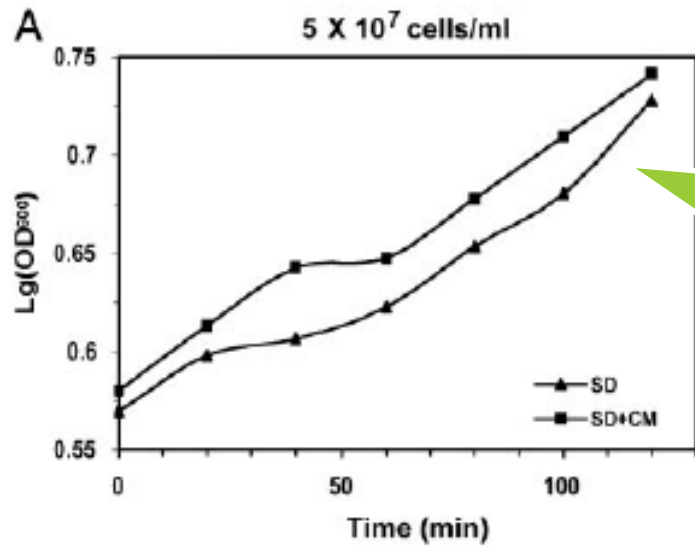
By Chen and colleagues(2004)

Long lag phase for diluted *C. albicans* inoculum

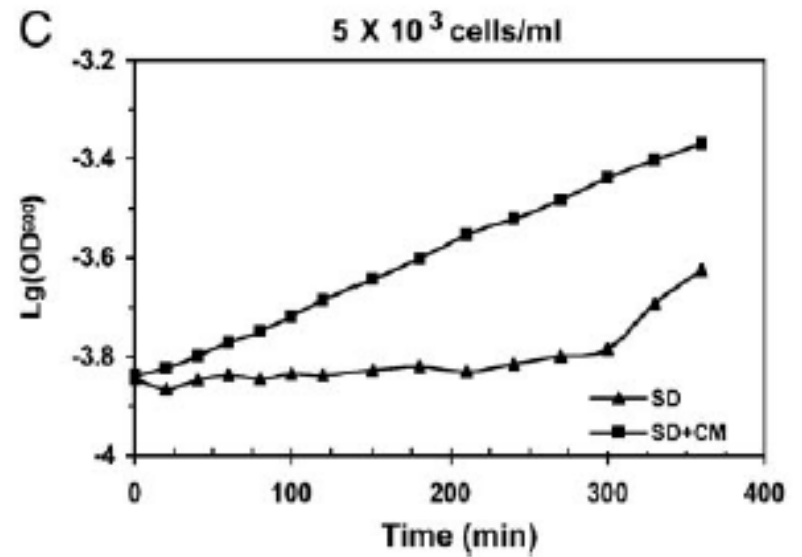
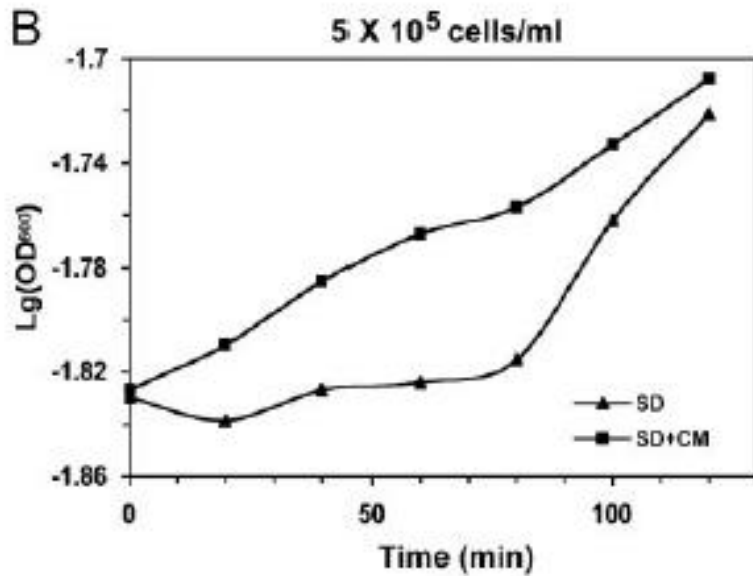
- More diluted, longer the lag phase
- + Spent media, lag phase shortened
- Active molecule: Tyrosol

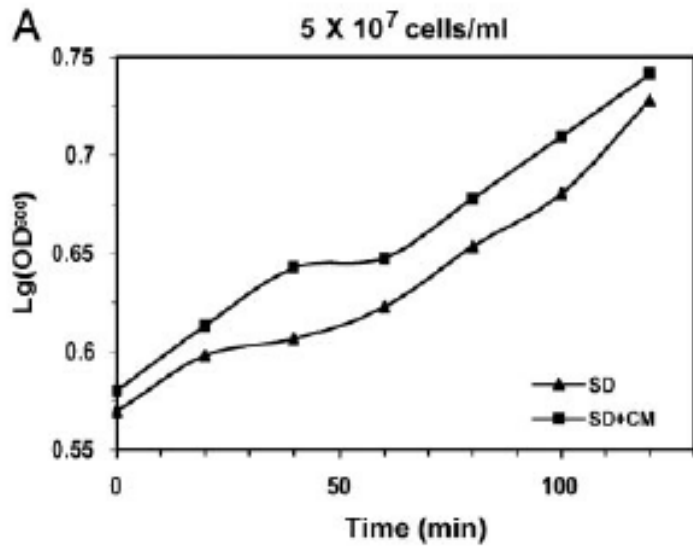


<https://upload.wikimedia.org/wikipedia/commons/a/a6/Tyrosol.png>

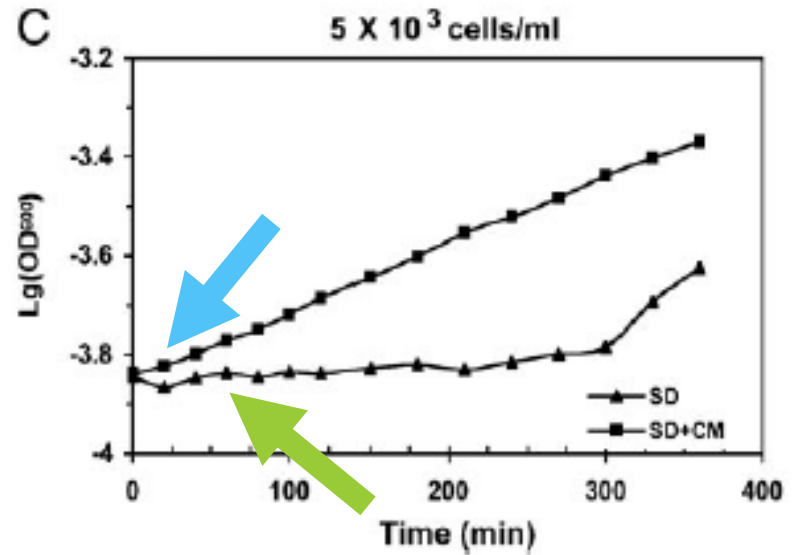
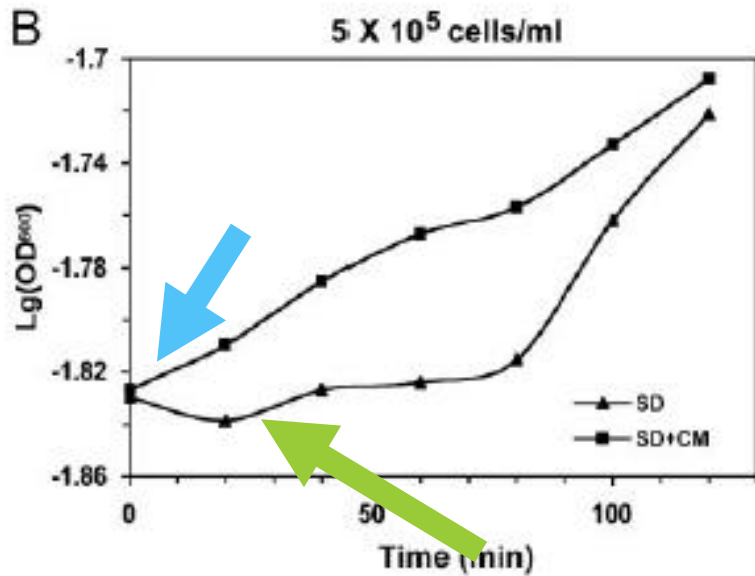


$5 \times 10^7$  cells/ml  
High inoculum concentration:  
No lag phase





5x10<sup>5</sup> & 5x10<sup>3</sup> cells/ml  
 Low inoculum concentration:  
 Lag phase presences



+ Conditional media: lag phase shortens

# Other properties of tyrosol

- Commercial tyrosol: same effect
- Production roughly proportional to cell density
- Transcriptional regulation of DNA-replication machinery and cell-cycle-control proteins
- Promote germ tube formation

# Other properties of tyrosol

- Production in biofilms > in planktonic cells
- Early and intermediate stages:  
Tyrosol's effect > Farnesol's effect
- Mature biofilm:  
Farnesol's effect & concentration > Tyrosol's  
→ May be linked to biofilm dispersal



# Examples of other possible QS system found in yeast

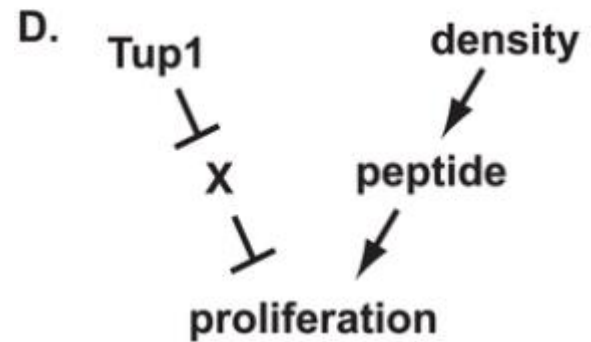
## *Saccharomyces cerevisiae*

- QSMs: Typtophol, Phenylethanol
  - Control filamentation in low nitrogen conditions

## *Cryptococcus neoformans*

- QS-like behavior in mutants of TUP1 of *C. neoformans*:  
No growth if [inoculum]  $< 10^3$  cells/ml
- + Conditional medium from high density culture  $\rightarrow$  growth rescued
- Active molecule: peptide

C. NH<sub>2</sub>-NFGAPGGAYPW-COOH



# Possible applications of yeast QSMs

## Clinical purposes:

- Farnesol from *C. albicans*
- Preventing biofilm on catheter and implant
- New type of antifungals or antibiotics
- Developing quorum sensing inhibitors

## Industrial purposes:

- Monitoring and controlling fermentation process
- Studying interactions between microbes

# Conclusion

- Quorum sensing
  - Modulation of gene expression
  - Cell-density-dependent manner
- Quorum sensing molecules in yeast:
  - Farnesol: Inhibit filamentation
  - Tyrosol: Shorten lag phase, promote germ tube formation
  - New ways to prevent and cure microbial infection

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